

Toxicological risk assessment of cyano blooms & cyanotoxins

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In cooperation with RAWAT consulting
for HYDAP

Kick-off meeting

Basic parameters used in risks evaluation of cyanotoxins:

- Biomass development trends
- Biomass density
- Cyanotoxins content
- Forecast parameters
 - Weather
 - Nutrient flow (external, internal)
 - Inoculum in sediments
 - Ecophysiology of populations

Are there any alternatives for cyanotoxins risks assessment?

- **General idea:**

CYANOBACTERIA – general parameters for cyanobacterial biomass (raw water)

- Biovolume
- Pigments composition

CYANOTOXINS (treated water)

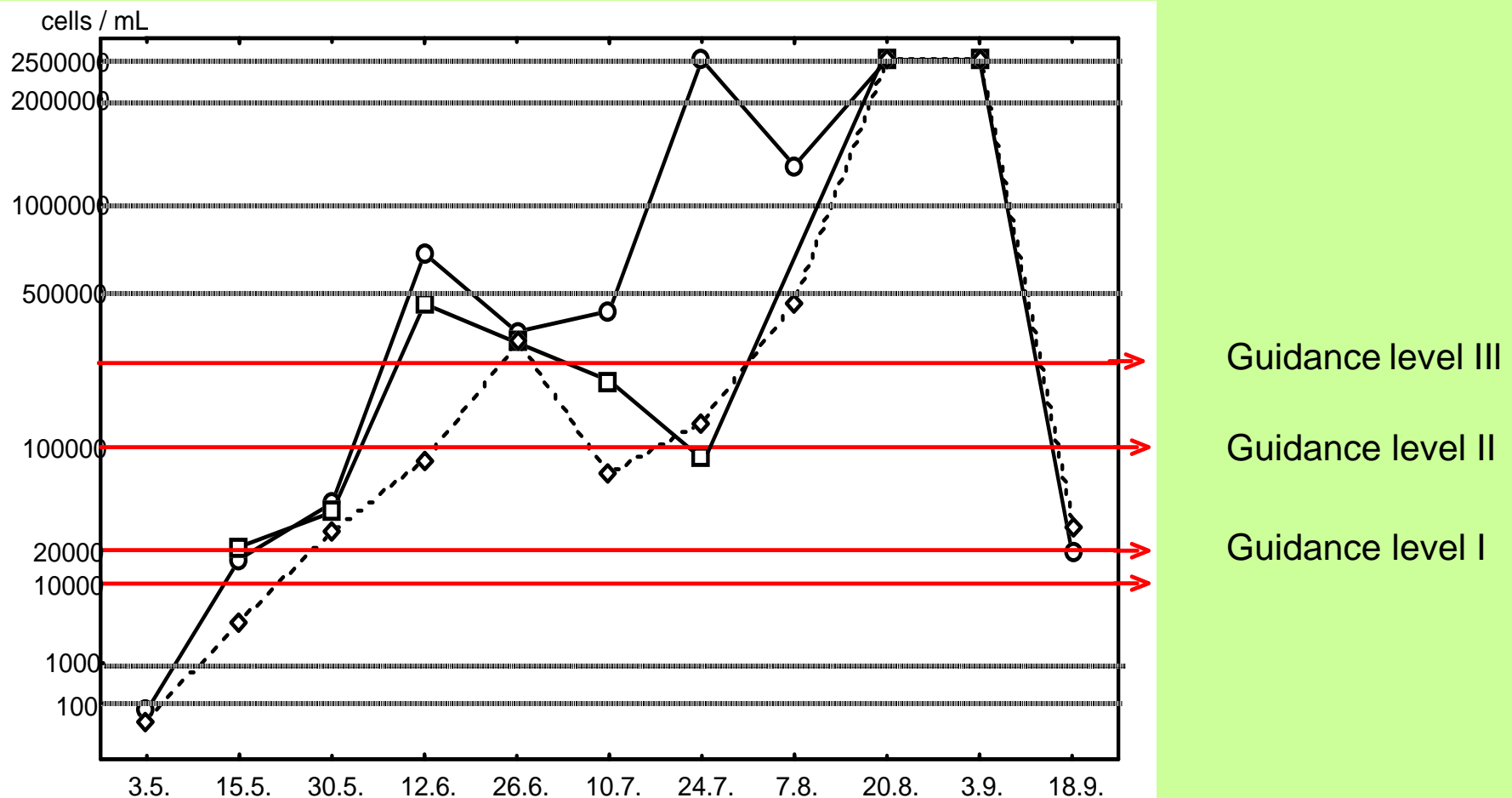
- Sensors for general toxicity
- Sensors for selected toxins

Cyanobacterial BIOMASS

- Cyanobacterial biomass is the key monitored parameter in surface waters
HOWEVER:
 - intercalibration study for MICROCYSTIS quantification (**cell counts**) by 37 routine laboratories give the variability unacceptable by GLP
- **Chlorophyll a** concentrations by ISO method and FluoroProbe fits well, but ISO method did not discriminate cyanobacteria from other phytoplankton
- **Microscopic determination and biovolume** give a good results, but it is time-consuming and need the experience
- COMBINATION OF METHODS IS NEEDED:
- **Fluoro Probe** and the dominant species determination seems to be a good compromise for water management

Dermal contact with cyanobacteria in Brno reservoir represents relatively high health risk during a substantial part of the summer season.

Concentrations of cyanobacteria in Brno reservoir 2007(cells / mL) and comparison with WHO safety "Guidance levels". Results from three localities sampled biweekly.

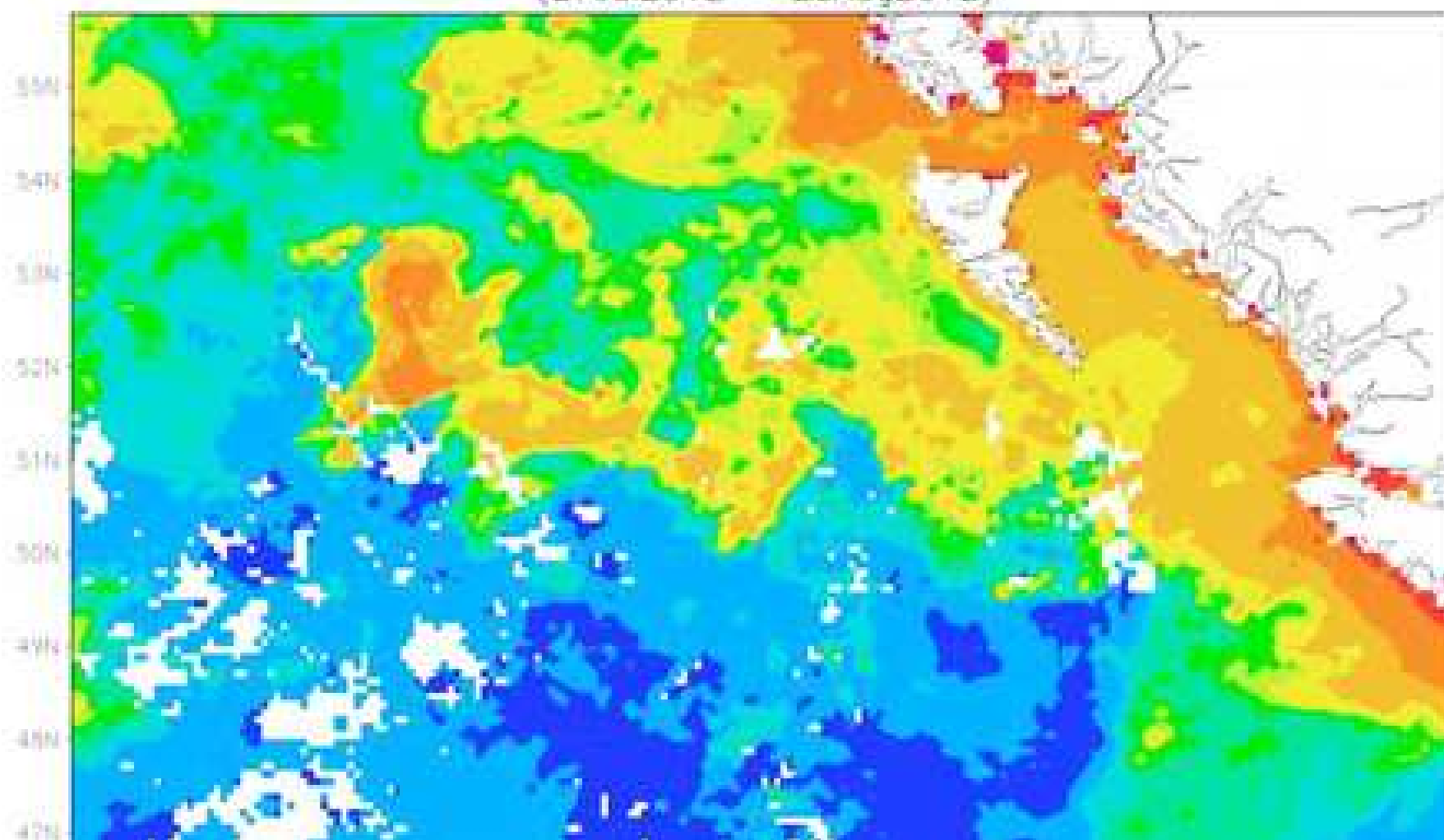




10 000 cells/ml

> 1 000 000 cells/ml

MA8D_CHLO_9km.CR Chlorophyll a concentration [mg/m³]
(27Jul2012 - 28Aug2012)

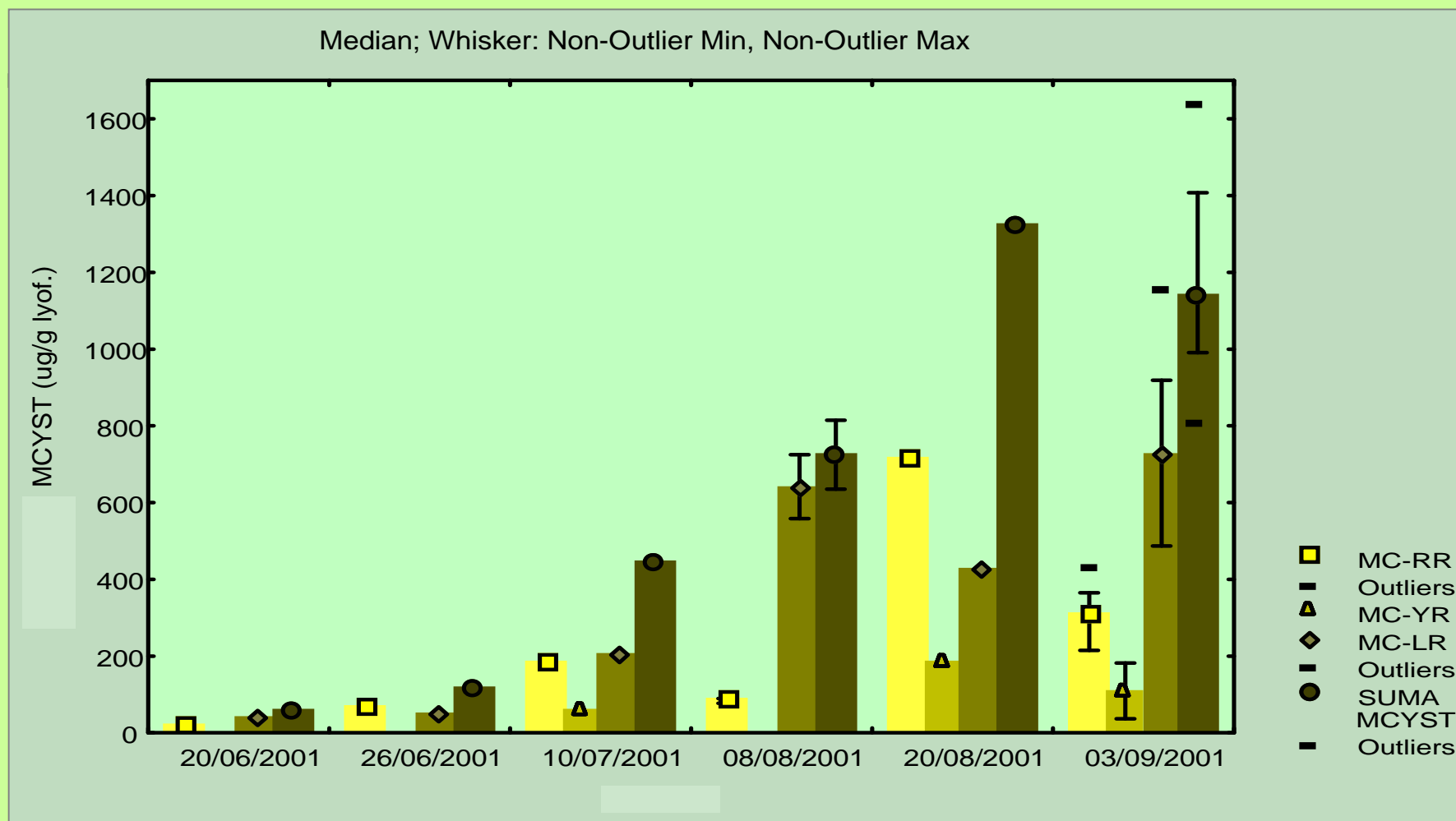


Intake of microcystins during swimming represents highly significant risks in the Brno reservoir both for children and adults.

Particularly, during August high HI values (up to 40 for children) were observed. HI > 1 for children were observed at 82% of localities during August 2001.

date	water conc. of MCYST ($\mu\text{g.l}^{-1}$)	children (25 kg, intake of 80 ml water.h ⁻¹)				adults (70 kg, intake 50 ml.h ⁻¹)			
		regular swim. (2,6 h.d ⁻¹)		weekend swim. (5 h.d ⁻¹)		regular (2,6 h.d ⁻¹)		weekend (5 h.d ⁻¹)	
		intake MCYST ($\mu\text{g.kg}^{-1}$ b.w./d)	HI	intake MCYST ($\mu\text{g.kg}^{-1}$ b.w./d)	HI	intake MCYST ($\mu\text{g.kg}^{-1}$ b.w./d)	HI	intake MCYST ($\mu\text{g.kg}^{-1}$ b.w./d)	HI
26.6.	1,46	0,0122	0,30	0,0234	0,58	0,0027	0,07	0,0052	0,13
10.7.	5,86	0,0488	1,22	0,0938	2,35	0,0109	0,27	0,0209	0,52
7.8.	48,19	0,4010	10,02	0,7711	19,28	0,0895	2,24	0,1721	4,30
20.8.	116,18	0,9666	24,17	1,8589	46,47	0,2158	5,39	0,4149	10,37
3.9.	99,76	0,8300	20,75	1,5961	39,90	0,1853	4,63	0,3563	8,91

Fig. 1: Variations of three dominating microcystin variants in the cyanobacterial water blooms collected from Brno reservoir during the 2001 season. Median values (+/- maximum/minimum).



Brno-reservoir
September 2001

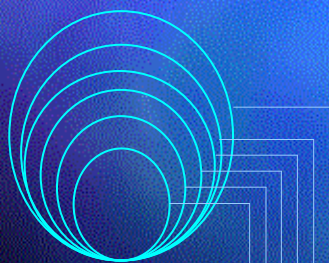
3. 9. 2001

△ MC-LR

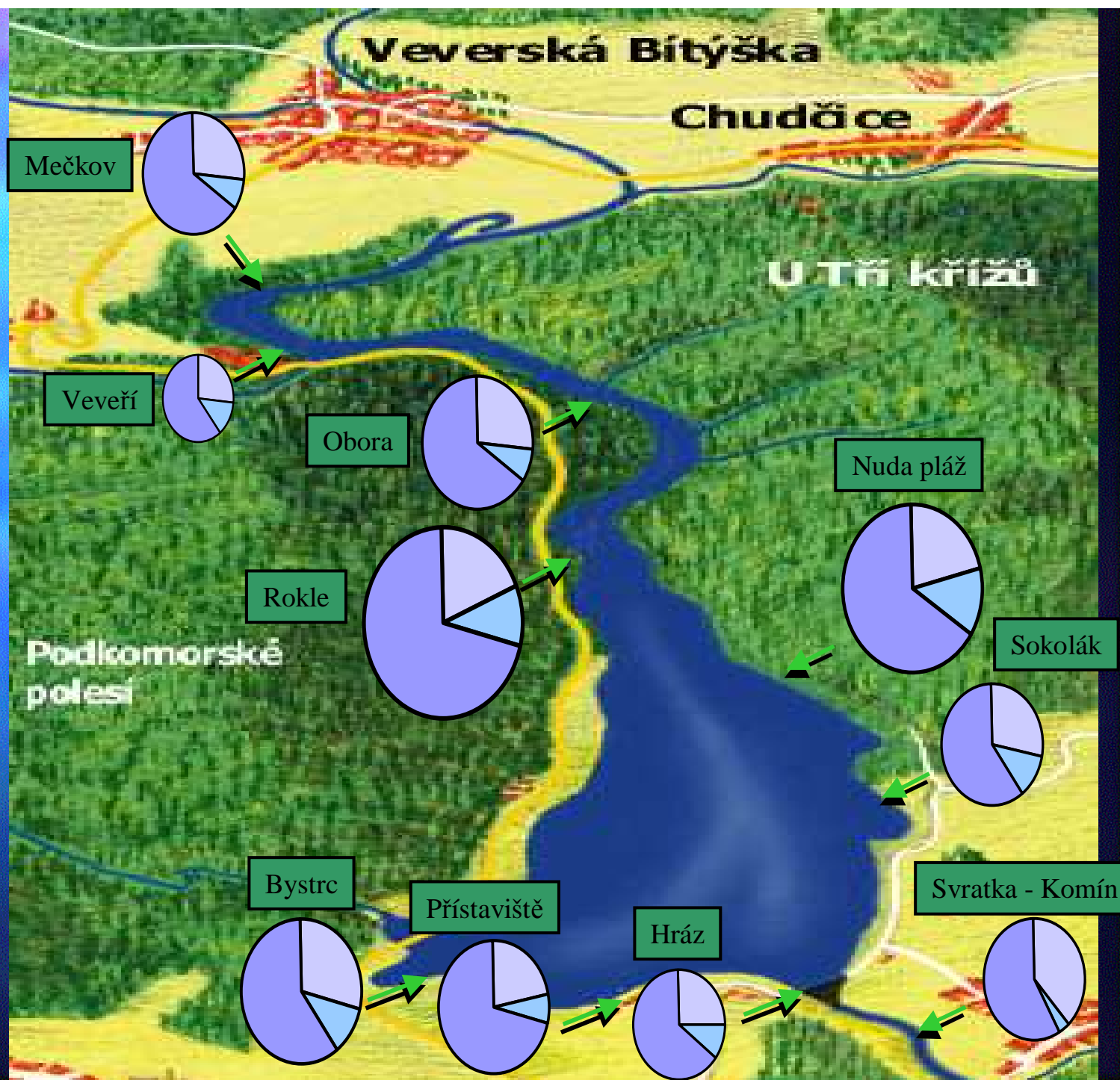
△ MC-YR

△ MC-RR

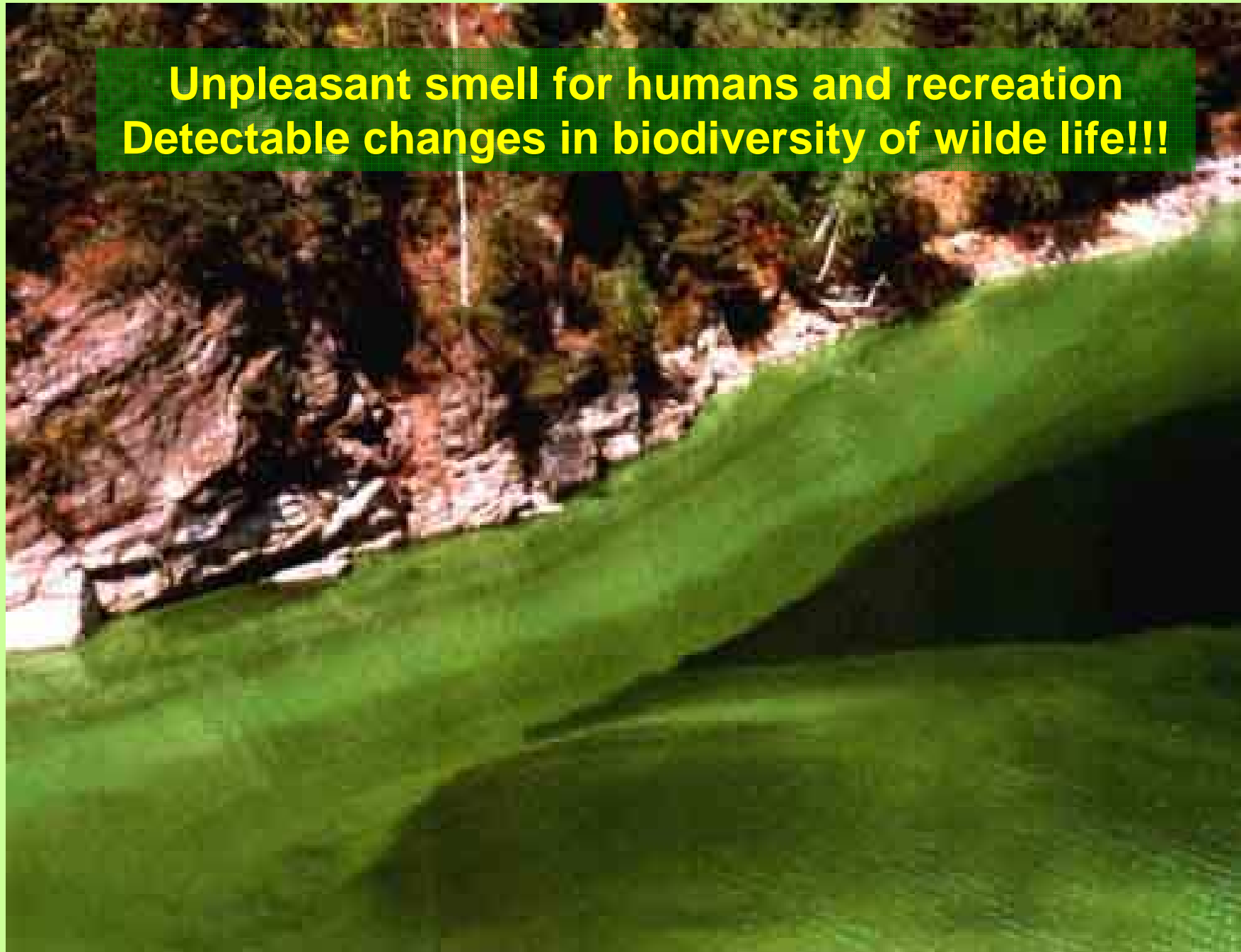
Koncentration
MCYST (total)



< 891 $\mu\text{g.g}^{-1}$ lyof.
891 - 974 $\mu\text{g.g}^{-1}$ lyof.
974 - 1057 $\mu\text{g.g}^{-1}$ lyof.
1057 - 1140 $\mu\text{g.g}^{-1}$ lyof.
1140 - 1223 $\mu\text{g.g}^{-1}$ lyof.
1223 - 1306 $\mu\text{g.g}^{-1}$ lyof.
1306 - 1389 $\mu\text{g.g}^{-1}$ lyof.
1389 - 1472 $\mu\text{g.g}^{-1}$ lyof.
1472 - 1555 $\mu\text{g.g}^{-1}$ lyof.
>1555 $\mu\text{g.g}^{-1}$ lyof.



**Unpleasant smell for humans and recreation
Detectable changes in biodiversity of wildlife!!!**



Trends in the risk evaluation

- Toxicity identification require special expertise and it is expensive
- All cyanobacteria produce toxins
- The most practical way is to quantify cyanobacterial biomass (pigments or cells by microscope, or special device like Fluoro Probe or CCD kinetic fluorescence immaging

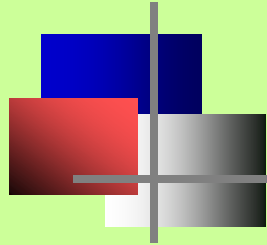
Methodological approaches- prediction

- Fluorescence microscopy and probes
- Laboratory cultivation
- CCD camera with FLIA was proved as powerful tool for the study of annual cycle of colony formation and ecophysiology of *Microcystis*
- meticulous care and long time under microscope was spend to build up base for
 - next comparative study of seasons
 - other localities
 - European countries
 - other species

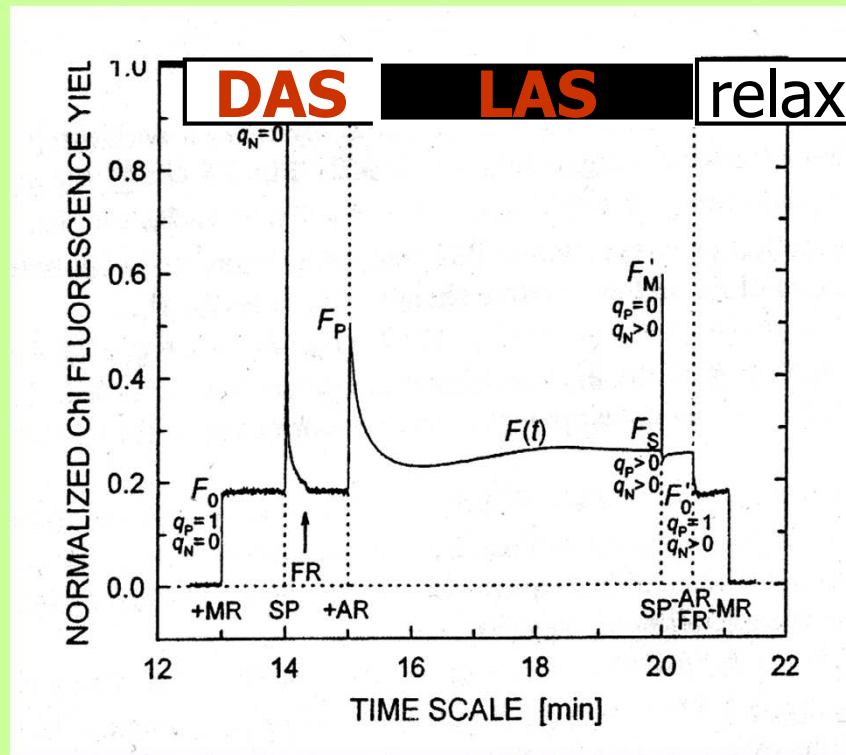
Principle

- Light-induced PSII fluorescence kinetic
 - Shows photosystem associated electron transport chain processes
 - Fluorescence yield is connected to chlorophyll and redox state of quinons as well as to their binding sites disposition in photosystem
- => Detection of effects at base level of energy metabolism of autotrophic cell

Chlorophyll fluorescence kinetic record

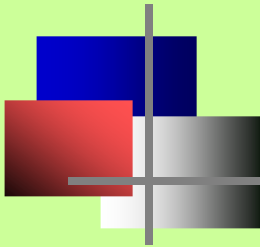


1. Dark Adapted State
2. Light Adapted State



Several basic terms are measured: F_0 F_m F_p
 F_s $F_{m'}$ $F_{0'}$

Fast and slow kinetic could be used



Purpose of evaluation

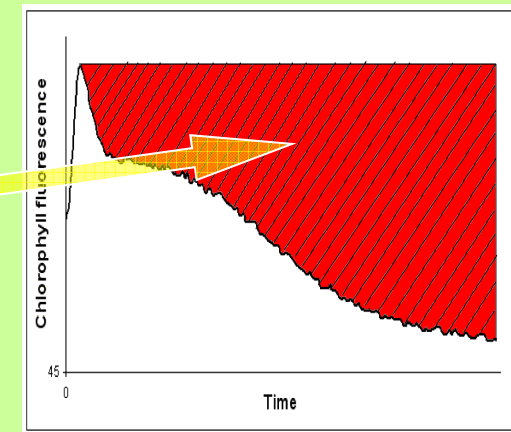
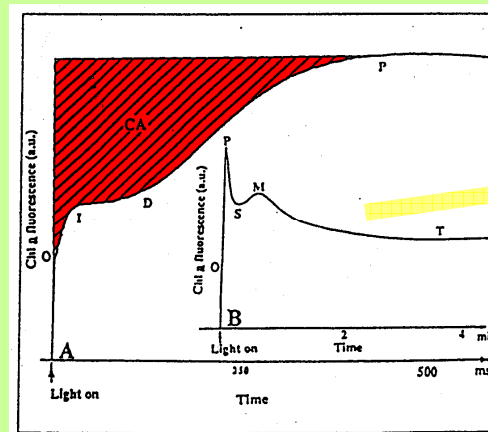
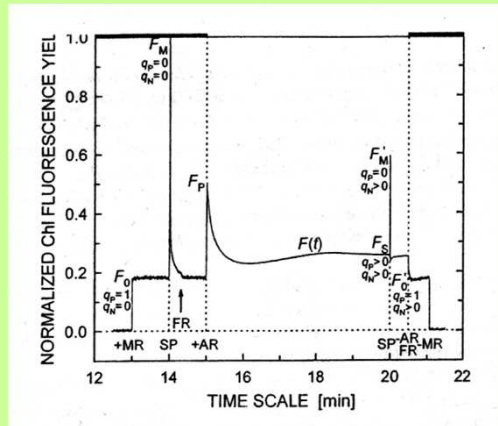
Fluorescence parameters + fluorimeters

- specific mode of action
- studies of physiology or processes in photosynthesis

Complementary area + fluorescence imaging

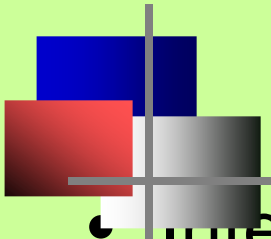
- Screening bioassays
- Fluorescence imaging of heterogeneous and micro scale objects

Evaluation of the processes



1] by the terms and fluorescence parameters of the slow kinetics

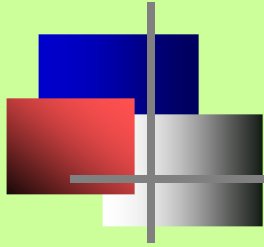
2] by calculating definite integral residual (complementary area) from fast and slow kinetic records of fluorescence



Benefits of evaluation by slow complementary area

- Integrates several modes of action (ETS, proton pump, processes on membranes, ATP synthesis, Calvin cycle)
- Simple setup, simple interpretation
- Increase sensitivity for CCD fluorescence imaging systems
- Give the chance to observe heterogeneous object with low chlorophyll content
- Shows small variability in replicates

Instruments



- Our instrumentation is based on fluorescence imaging systems (PSI, Brno, Czech Republic) with CCD camera detection

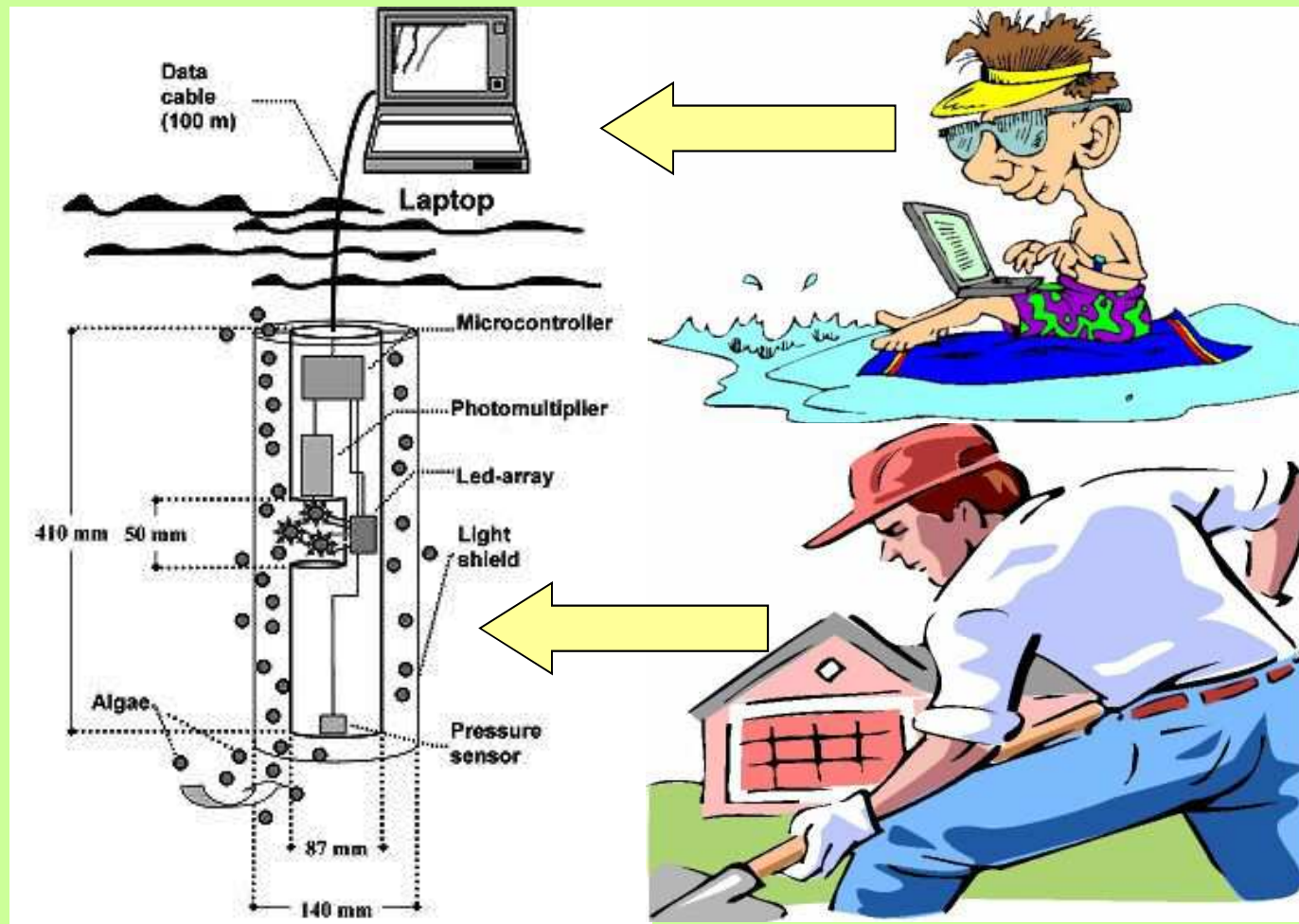
Photosynthetic pigments

<i>group</i>	<i>pigments</i>	<i>Excit. (nm)</i>	<i>Emis. (nm)</i>
Chlorophyta	Chlorophyll-b	480	685
Chromophyta Dinophyta	Chlorophyll-c Karotenoidy	460 525	685
Cyanobacteria	phycocyanin	610	650-685
Cryptophyta	phycoerythrin Chlorophyll-c	550 460	685

Fluoro Probe



FluoroProbe



03 Sokolak

Data | Graphics 1 | Graphics 2 | Common parameters | Parameters of measurement | Parameters of fit

time	depth [m]	temp. [°C]	algae [µg/l]				total [µg/l]	transm. [%]	int. temp.	
			Green	Bluegr.	Diatom.	Crypto.			[°C]	[°C]
06:55:59	0,03	20,9	0,00	0,20	0,00	0,00	0,20	56,05	24,2	22,6
06:56:02	0,09	21,1	1,21	1,63	0,00	0,00	2,83	54,86	24,3	22,6
06:56:04	0,23	21,3	5,92	8,89	0,00	1,17	15,97	65,55	24,3	22,6
06:56:06	0,27	21,4	5,36	7,81	0,00	1,67	14,84	65,78	24,3	22,7
06:56:09	0,39	21,4	5,65	7,52	0,00	1,58	14,74	66,08	24,3	22,7
06:56:11	0,50	21,4	5,96	7,45	0,00	1,69	15,11	65,78	24,3	22,7
06:56:14	0,67	21,4	5,66	8,31	0,00	1,68	15,65	66,07	24,3	22,7
06:56:16	0,81	21,4	5,50	7,78	0,00	1,91	15,19	66,00	24,3	22,7
06:56:18	1,02	21,4	5,63	7,70	0,00	1,98	15,31	65,82	24,3	22,7
06:56:21	1,21	21,4	5,78	7,55	0,00	1,66	14,99	66,22	24,3	22,7
06:56:23	1,58	21,4	5,88	6,32	0,00	1,69	13,88	66,28	24,3	22,8
06:56:25	2,08	21,4	6,23	5,56	0,00	1,17	12,97	66,50	24,3	22,8

results: ☐ (raw data)

First

Minus 12

Prev

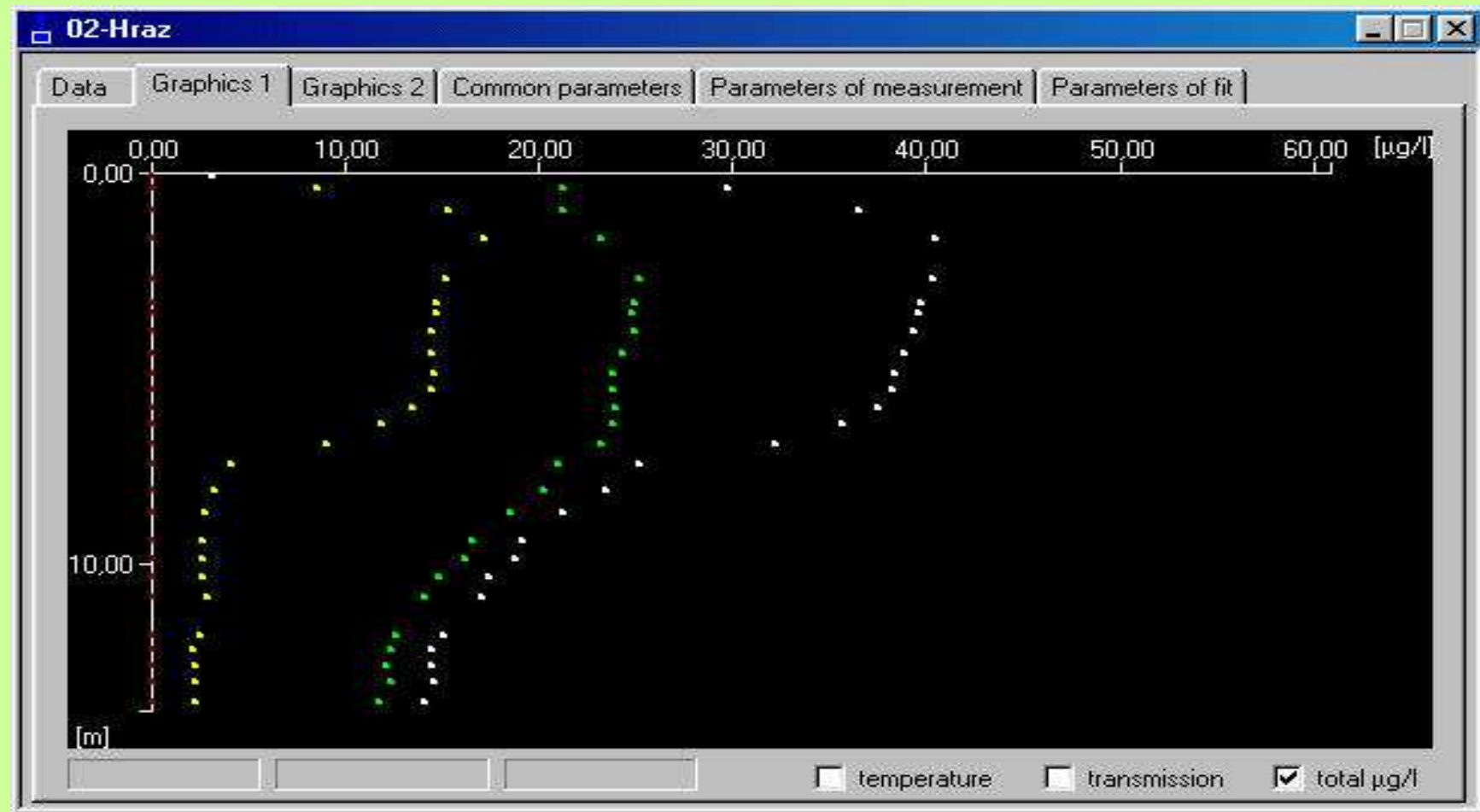
Next


Plus 12

Last

Fullscreen

FluoroProbe - graph





**Cyanotoxins and
cyanobacterial blooms
management represents a great
potential for the remote sensing
application!!!!**

THANK YOU

Expected results

- Joint effort for validation and calibration of data measured by in-lake and aircraft spectra
- Evaluation of RS tools potential for risks assessment of cyanobacterial blooms (dynamic of biomass movement)
- Cross discussion - RS tools potential for water quality assessment (soil erosion, nutrients pollution, use of new devices, spectra combination etc.).